

## Reference Data

**Discrimination of Structural Isomers of *N*-Methylated and *N*-*tert*-Butylated Tetrazoles by  $^{13}\text{C}$  and  $^{15}\text{N}$  NMR. Long-Range  $^{15}\text{N}$ ,  $^1\text{H}$  Coupling Constants**

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Natural abundance  $^{13}\text{C}$  and  $^{15}\text{N}$  NMR studies were carried out on a series of *N*-methylated and *N*-*tert*-butylated tetrazoles. Long-range  $^{13}\text{C}$ ,  $^1\text{H}$  and  $^{15}\text{N}$ ,  $^1\text{H}$  NMR coupling constants together with  $^1J(^{15}\text{N}, ^{13}\text{C})$  allowed unequivocal assignments of  $^{13}\text{C}$  and  $^{15}\text{N}$  chemical shifts for isomeric 1,5- and 2,5-disubstituted tetrazoles. © 1997 by John Wiley & Sons, Ltd.

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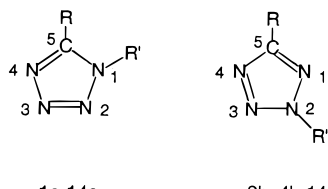
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## INTRODUCTION

The identification of structural isomers of alkylated tetrazole and its 5-substituted derivatives has been the focus of many investigations, including  $^1\text{H}$ ,  $^{13}\text{C}$  and  $^{15}\text{N}$  NMR methods.<sup>1</sup> In previous  $^{15}\text{N}$  NMR studies the chemical shift assignments were not straightforward and attempts based upon MO electron density calculations led to contradictory results.<sup>1,2</sup> Long-range  $^{13}\text{C}$ ,  $^1\text{H}$  and  $^{15}\text{N}$ ,  $^1\text{H}$  coupling constants could provide more information in terms of unambiguous identification and structure elucidation of isomeric tetrazoles. Accordingly, we performed a detailed  $^{13}\text{C}$  and  $^{15}\text{N}$  NMR study of a series of methylated (1–7) and *tert*-butylated (8–14) tetrazoles. In the case of 1–7 we prepared  $^{13}\text{C}$  labelled *N*-methylated compounds, which allowed us also to observe  $^{13}\text{C}$ ,  $^{13}\text{C}$  and  $^{15}\text{N}$ ,  $^{13}\text{C}$  coupling constants.

## RESULTS AND DISCUSSION

The  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts for 1–14 are given in Table 1. In the case of 5-*H*-tetrazole and 5-*tert*-butyltetrazole we obtained only the 1-methyl isomers 1a and 3a. For 2 and 4–7 the methyl protons of the 1-isomers are slightly more shielded by 0.15–0.53 ppm compared with their 2-substituted analogues, but for the *tert*-butyl protons of the isomers of 8–14 no such consistency was observed.

		Compound	R'	R
		1	Me	H
		2	Me	Me
		3	Me	<i>t</i> -Bu
		4	Me	cyclopropyl
		5	Me	Ph
		6	Me	$\text{CH}_2\text{Ph}$
		7	Me	$(\text{CH}_2)_2\text{Ph}$
		8	<i>t</i> -Bu	H
		9	<i>t</i> -Bu	Me
		10	<i>t</i> -Bu	<i>t</i> -Bu
		11	<i>t</i> -Bu	cyclopropyl
		12	<i>t</i> -Bu	Ph
		13	<i>t</i> -Bu	$\text{CH}_2\text{Ph}$
		14	<i>t</i> -Bu	$(\text{CH}_2)_2\text{Ph}$

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Carbon chemical shift assignments were made on the basis of signal intensities, analysis of  $^{13}\text{C}$ ,  $^1\text{H}$  NMR coupling patterns, including selective proton decoupled experiments, and literature data.<sup>4,5</sup> For all pairs of isomers, the largest difference in carbon chemical shifts was observed for C-5 (Table 1), which for the 2,5-isomer resonates 9.20–12.20 ppm to higher frequency relative to the 1,5-isomer. The isomeric methylated tetrazoles 2 and 4–7 could be distinguished on the basis of long-range  $^{13}\text{C}$ ,  $^1\text{H}$  NMR coupling constants involving C-5 and the protons of the methyl substituent at N-1 or N-2 (Table 2), as in the case of N-1 substitution we observed  $^3J_{\text{C-5, HMe}}$  (2.1–2.8 Hz). For the 2-isomers  $^4J_{\text{C-5, HMe}}$  was detected only in 5b and was much smaller than  $^3J_{\text{C-5, HMe}}$  in 5a. The values of the chemical shifts,  $^1J_{\text{CH}}$  coupling constants and substitution chemical shifts of the aromatic carbons for 5a,b and 12a,b are given in Table 3. The largest differences for pairs of isomers are observed for the chemical shifts of C-1' and C-2'. The  $^1J_{\text{CH}}$  coupling constants in 5a,b and 12a,b (160.9–163.2 Hz) are slightly greater than those for 6a,b, 7a,b, 13a,b and 14a,b (147–160 Hz), in which the tetrazole and phenyl rings are separated by one or two methylene groups. Substituent carbons attached to N-1 are more shielded than those attached to N-2 atoms in the 2, 4–7 and 8–14 series.

Absolute values of  $^1J_{\text{CN}}$  coupling constants for 1–7 are given in Table 4. The signs of the constants are presumably negative<sup>6</sup> and the absolute values of  $^1J_{\text{C, N-1}}$  are smaller than the corresponding  $^1J_{\text{C, N-2}}$  by 1.9–2.1 Hz. Table 4 also contains the magnitudes of the  $^{13}\text{C}$ ,  $^{13}\text{C}$  coupling constants in 1–7. Vicinal transoid couplings in 2b and 4b–7b are slightly larger than or equal to the geminal couplings in 1a–7a, and the magnitudes resemble the couplings found in aromatic compounds with aliphatic substituents.<sup>7</sup>

The  $^{15}\text{N}$  NMR chemical shifts and the absolute values of  $^{15}\text{N}$ ,  $^1\text{H}$  NMR coupling constants are presented in Table 5. Our assignments were based mostly on the analysis of  $^{15}\text{N}$ ,  $^1\text{H}$  NMR coupling patterns with the use of selective proton decoupling where necessary. In previous reports the assignments were connected with the assumption that N-1 and N-4 in general resonate at lower frequency and N-2 and N-3 resonate at higher frequency in tetrazoles and that alkyl substitution results in significant shifting of the pyrrole-type nitrogen resonance to lower frequency. However, relative positions of the signals were not established unambiguously. For an extensive series of tetrazoles it was shown that  $^{15}\text{N}$  NMR chemical shifts correlate well with the electron densities at the nitrogen atoms,<sup>1</sup> but in the case of 2,5-dimethyltetrazole (2b) different MO methods resulted in contradictory assignments concerning the relative positions of the N-3 and N-4 signals.<sup>1,2</sup> In the proton-coupled spectrum of 2b both the N-3 and N-4 signals appear as quartets with  $^{15}\text{N}$ ,  $^1\text{H}$  coupling constants that are close in magnitude (Table 5), supposedly coupled to the protons of *N*-Me and *C*-Me groups, respectively. Irradiation of the *C*-Me protons at 2.11 ppm affected the lower frequency N signal (–48.00 ppm), turning it into a singlet, whereas for the lower frequency N signal the splitting was preserved. This agrees with the results reported in Ref. 2. Unambiguous  $^{15}\text{N}$  NMR chemical shift assignments in tetrazoles require coupling pattern analysis and cannot be based upon the MO calculations of electron densities alone. It should be noted that for 9b, 10b, 12b and 14b we detected four-bond  $^{15}\text{N}$ ,  $^1\text{H}$  couplings between *tert*-butyl protons and N-3, which were also confirmed by selective proton decoupling. The  $^{15}\text{N}$  NMR chemical shifts become increasingly more negative in the order N-3 < N-2 < N-4 < N-1 for 1,5- and N-3 < N-4 < N-1 < N-2 for 2,5-disubstituted tetrazoles.

$^1\text{H}$  and  $^{13}\text{C}$  chemical shifts can be used to differentiate between isomeric 1,5- and 2,5-disubstituted tetrazoles only in special circumstances. In contrast, the application of  $^{15}\text{N}$  chemical shifts for this purpose is general. This work demonstrates that the  $^{13}\text{C}$  and  $^{15}\text{N}$

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**Table 1.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts ( $\delta$ , ppm) for compounds 1–14, with proton chemical shifts (Hz) in parentheses<sup>a</sup>

Compound	Me <sub>R</sub> <sup>c</sup>	C <sub>f-Bu</sub>	C-5	R
<b>1a</b> <sup>b</sup>	33.80 (3.9)	—	143.20 (8.7)	—
<b>2a</b>	32.72 (3.7)	—	151.42	Me: 7.83 (2.2)
<b>2b</b>	38.46 (3.9)	—	162.00	Me: 9.86 (2.1)
<b>3a</b>	35.69 (4.1)	—	160.76	Me <sub>f-Bu</sub> : 28.22 (1.4); C <sub>f-Bu</sub> : 30.85
<b>4a</b>	32.48 (3.8)	—	156.55	Cyclopropyl: 2.84(CH); 7.71(CH <sub>2</sub> ) (1.6, 0.7–0.9)
<b>4b</b>	38.43 (3.9)	—	167.80	Cyclopropyl: 5.52(CH); 7.49(CH <sub>2</sub> ) (1.6, 0.7–0.9)
<b>5a</b>	34.03 (4.0)	—	153.18	Ph: 122.75; 127.52; 128.07; 130.05 (7.3–7.6)
<b>5b</b>	38.35 (4.2)	—	163.76	Ph: 125.57; 126.43; 127.86; 129.18 (7.3–7.6, 8.1)
<b>6a</b>	32.50 (3.7)	—	153.08	CH: 28.36 (4.1)
				Ph: 126.35; 127.57; 127.89; 133.14 (7.0–7.2)
<b>6b</b>	38.12 (4.1)	—	164.37	CH <sub>2</sub> : 30.53 (4.0)
				Ph: 125.72; 127.53; 127.74; 135.95 (7.0–7.2)
<b>7a</b>	32.20 (3.3)	—	153.88	CH <sub>2</sub> CH <sub>2</sub> : 24.28; 32.53 (2.2, 2.7)
				Ph: 125.94; 127.62; 127.92; 138.75 (6.7–6.9)
<b>7b</b>	38.37 (3.8)	—	165.14	CH <sub>2</sub> CH <sub>2</sub> : 26.36; 33.17 (2.2, 2.7)
				Ph: 125.45; 127.54; 127.67; 139.75 (6.7–6.9)
<b>8a</b> <sup>c</sup>	29.35 (1.6)	59.40	139.81 (8.6)	—
<b>8b</b> <sup>b</sup>	28.98 (1.7)	63.66	152.02 (8.4)	—
<b>9a</b>	28.89 (1.7)	60.03	150.20	Me: 11.43 (2.7)
<b>9b</b>	28.62 (1.7)	62.69	161.40	Me: 10.24 (2.5)
<b>10a</b>	30.32 (1.7)	62.25	160.93	Me <sub>f-Bu</sub> : 30.62 (1.5); C <sub>f-Bu</sub> : 32.34
<b>10b</b>	28.60 (1.6)	62.61	173.11	Me <sub>f-Bu</sub> : 28.92 (1.3); C <sub>f-Bu</sub> : 30.81
<b>11a</b>	28.69 (1.4)	59.70	155.41	Cyclopropyl: 4.27(CH); 8.51(CH <sub>2</sub> ) (0.6–0.8, 1.8)
<b>11b</b>	28.37 (1.3)	62.41	166.96	Cyclopropyl: 5.79(CH); 7.44(CH <sub>2</sub> ) (0.6–0.8, 1.8)
<b>12a</b>	29.93 (1.6)	61.45	153.38	Ph: 126.84; 129.30; 128.03; 130.07 (7.2–7.3)
<b>12b</b>	28.85 (1.8)	63.30	163.93	Ph: 126.27; 127.44; 128.31; 129.55 (7.2–7.3, 8.1)
<b>13a</b>	28.82 (1.4)	60.26	151.87	CH <sub>2</sub> : 30.42 (4.22)
				Ph: 126.41; 127.56; 128.01; 134.50 (6.9–7.1)
<b>13b</b>	28.38 (1.4)	62.51	163.82	CH <sub>2</sub> : 30.99 (4.00)
				Ph: 125.84; 127.70; 127.91; 136.29 (6.9–7.1)
<b>14a</b>	29.17 (1.6)	60.06	153.24	CH <sub>2</sub> CH <sub>2</sub> : 27.74, 33.26 (3.0–3.2)
				Ph: 126.28; 127.95; 128.34; 139.72 (7.0–7.2)
<b>14b</b>	28.82 (1.6)	62.83	164.82	CH <sub>2</sub> CH <sub>2</sub> : 27.02, 33.78 (3.0–3.2)
				Ph: 125.83; 128.04; 128.04; 140.38 (7.0–7.2)

<sup>a</sup> Chemical shifts and coupling constants are accurate to  $\pm 0.01$  ppm and  $\pm 0.1$  Hz, respectively.

<sup>b</sup>  $^1J_{\text{C-5, H-5}} = 217.3$  Hz.

<sup>c</sup>  $^1J_{\text{C-5, H-5}} = 214.1$  Hz for **8a** and 211.9 Hz for **8b**.

NMR chemical shifts for each isomer can be unequivocally assigned by consideration of the magnitudes of  $^{15}\text{N}$ ,  $^1\text{H}$ ,  $^{13}\text{C}$ ,  $^1\text{H}$  and  $^{15}\text{N}$ ,  $^{13}\text{C}$  NMR coupling constants.

## EXPERIMENTAL

Methylated tetrazoles 1–7 were synthesized by alkylation of 5-substituted tetrazoles with  $^{13}\text{C}$ -labeled iodomethane.<sup>8</sup> *tert*-Butylation of 5-substituted tetrazoles was performed as described in the literature.<sup>9</sup>  $^1\text{H}$  and  $^{13}\text{C}\{^1\text{H}\}$  NMR spectra of 50–100 mg ml<sup>-1</sup> solutions of 1–14 in CDCl<sub>3</sub> were recorded on a General Electric QE-300 NMR spectrometer, operating at 300.16 and 75.48 MHz, respectively, at ambient temperature, in 5 mm tubes, with TMS as internal standard.  $^{13}\text{C}$ ,  $^1\text{H}$  NMR coupling constants were directly measured from  $^{13}\text{C}$  NMR proton coupled spectra, recorded on a Varian Unity-plus 500 NMR spectrometer (125.70 MHz) in 5 mm tubes, with a 128 kHz spectral width, 3 s interpulse delay, 45° flip angle and zero-filling to

524 288 data points with final resolution of 0.06 Hz per point. Signs of the constants were not measured.

$^{15}\text{N}$  NMR spectra of 500–1000 mg ml<sup>-1</sup> of 1–14 in CDCl<sub>3</sub> were obtained on a Varian Unity-plus 500 NMR spectrometer, operating at 50.66 MHz, in 10 mm tubes, at ambient temperature. The  $^{15}\text{N}$  NMR chemical shifts were measured relative to external nitromethane. Spectral parameters were spectral width 36 kHz, 20 s interpulse delay, 45° flip angle, 128K data points and zero-filling with final resolution of 0.07 Hz per point.

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**Table 2.**  $^{13}\text{C}$ ,  $^1\text{H}$  coupling constants (Hz) of C-5 for compounds 1–7

Compound	$^2J(\text{C-5}, \text{H}_R)$	$^3J(\text{C-5}, \text{H}_R)$	$^3J(\text{C-5}, \text{H}_{Me})$
<b>1a</b>	—	—	2.2
<b>2a</b>	7.2	—	2.1
<b>2b</b>	7.2	—	—
<b>3a</b>	—	m <sup>b</sup>	m
<b>4a</b>	m	m	m
<b>4b</b>	m	m	m
<b>5a</b>	—	1.5	2.8
<b>5b<sup>a</sup></b>	—	4.1	—
<b>6a</b>	7.9	—	2.1
<b>6b</b>	8.0	—	—
<b>7a</b>	m	m	m
<b>7b</b>	m	m	m

<sup>a</sup>  $^4J_{\text{C-5}, \text{HMe}} = 0.75 \text{ Hz}$ .

<sup>b</sup> m = Multiplet.

**Table 4.** Absolute values of the  $^{13}\text{C}$ ,  $^1\text{H}$ ,  $^{13}\text{C}$ ,  $^{13}\text{C}$  and  $^{13}\text{C}$ ,  $^{15}\text{N}$  NMR coupling constants (Hz) of methyl carbons for compounds 1–7

Compound	$^1J_{\text{CH}}$	$^1J_{\text{CN}}$	$^2J_{\text{CC}}$	$^3J_{\text{CC}}$
<b>1a</b>	143.6	10.2	3.3	—
<b>2a</b>	142.8	11.4	3.1	—
<b>2b</b>	143.2	13.5	—	3.1
<b>3a</b>	142.8	10.4	2.3	—
<b>4a</b>	142.8	11.4	2.9	—
<b>4b</b>	143.3	13.5	—	3.1
<b>5a</b>	143.3	11.0	2.6	—
<b>5b</b>	143.7	13.0	—	3.0
<b>6a</b>	142.8	11.0	2.8	—
<b>6b</b>	143.4	13.1	—	3.1
<b>7a</b>	142.8	11.3	3.1	—
<b>7b</b>	143.4	13.2	—	3.1

**Table 3.**  $^{13}\text{C}$  NMR chemical shifts ( $\delta$ , ppm),  $^1J_{\text{CH}}$  coupling constants (Hz) and substitution chemical shifts (relative to benzene, 128.50 ppm) of the phenyl carbons of compounds 5a,b and 12a,b

	5a	5b	12a	12b
C-1'	122.75	126.43	126.84	127.44
C-2'	127.52	125.57	129.30	126.27
$^1J(\text{C-2}', \text{H-2}')$	161.7	163.2	162.0	161.5
C-3'	128.07	127.86	128.03	128.31
$^1J(\text{C-3}', \text{H-3}')$	162.2	161.4	162.2	160.9
C-4'	130.05	129.18	130.07	128.31
$^1J(\text{C-4}', \text{H-4}')$	161.9	163.0	161.9	160.9
<i>ipso</i> -	-5.75	-2.07	-1.66	-0.81
<i>ortho</i> -	-0.98	-2.93	+0.8	-2.23
<i>meta</i> -	-0.43	-0.64	-0.47	-0.19
<i>para</i> -	+1.55	+0.68	+1.57	+1.05

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**Table 5.**  $^{15}\text{N}$  NMR chemical shifts ( $\delta$ , ppm) and  $^{15}\text{N}, ^1\text{H}$  NMR coupling constants (Hz) for compounds 1–14

Compound	N-1	N-2	N-3	N-4
<b>1a</b>	–151.35 d, $^2J = 9.3$ q, $^2J = 2.0$	–8.29 m	14.29 d, $^3J = 3.0$	–48.64 d, $^2J = 12.1$
<b>2a</b>	–153.67 m	–7.49 q, $^3J = 1.9$	10.19 s	–52.06 q, $^3J = 1.9$
<b>2b</b>	–75.81 m	–103.62 q, $^2J = 2.3$	2.03 q, $^3J = 1.5$	–48.00 q, $^3J = 1.1$
<b>3a</b>	–156.81 m	–3.02 q, $^3J = 1.7$	8.03 s	–50.90 s
<b>4a</b>	–154.22 q, $^2J = 1.8$	–7.68 q, $^3J = 1.4$	9.18 s	–58.88 d, $^3J = 2.1$
<b>4b</b>	–81.00 m	–105.01 q, $^2J = 2.2$	1.23 q, $^3J = 1.8$	–53.53 d, $^3J = 1.3$
<b>5a</b>	–156.19 q, $^2J = 1.9$	–5.26 q, $^3J = 1.8$	11.39 s	–50.36 s
<b>5b</b>	–79.49 q, $^3J = 1.7$	–102.08 q, $^2J = 2.3$	3.15 q, $^3J = 1.6$	–51.83 s
<b>6a</b>	–151.15 q, $^2J = 2.2$	–2.41 q, $^3J = 2.2$	14.63 s	–44.11 m
<b>6b</b>	–71.79 q, $^3J = 1.7$	–99.93 q, $^2J = 2.2$	5.97 q, $^3J = 1.1$	–46.22 m
<b>7a</b>	–153.67 m	–7.45 q, $^3J = 1.7$	10.94 s	–51.39 t, $^3J = 1.9$
<b>7b</b>	–75.59 m	–103.71 m	2.31 q, $^3J = 1.2$	–48.01 t, $^3J = 1.4$
<b>8a</b>	–117.43 m	–11.36 d, $^3J = 1.1$	13.77 d, $^3J = 3.4$	–50.39 m
<b>8b</b>	–76.57 d, $^2J = 15.3$	–71.52 m	–2.18 s	–48.91 d, $^2J = 12.6$
<b>9a</b>	–125.00 m	–7.55 s	5.54 s	–48.62 q, $^3J = 2.2$
<b>9b</b>	–80.74 q, $^3J = 2.3$	–73.20 dec, $^3J = 2.3$	–1.52 dec, $^4J = 0.8$	–51.14 q, $^3J = 1.9$
<b>10a</b>	–124.18 m	–2.17 s	0.32 s	–46.06 s
<b>10b</b>	–82.82 s	–74.80 dec, $^3J = 2.5$	–2.64 dec, $^4J = 0.6$	–54.01 s
<b>11a</b>	–124.42 m	–8.03 s	3.34 s	–55.21 d, $^3J = 2.2$
<b>11b</b>	–85.65 d, $^3J = 1.2$	–74.67 dec, $^3J = 2.6$	–2.46 s	–57.28 d, $^3J = 1.7$
<b>12a</b>	–122.51 m	–2.87 s	6.73 s	–44.89 s
<b>12b</b>	–84.06 s	–71.92 dec, $^3J = 1.7$	–0.60 dec, $^4J = 0.7$	–54.51 s
<b>13a</b>	–124.70 m	–6.49 s	7.14 s	–44.75 t, $^3J = 1.8$
<b>13b</b>	–79.71 t, $^3J = 2.2$	–72.54 m, $^3J = 2.3$	–0.60 s	–49.71 t, $^3J = 3.0$
<b>14a</b>	–125.17 m	–8.38 s	5.83 s	–47.87 t, $^3J = 0.6$
<b>14b</b>	–80.58 t, $^3J = 1.2$	–73.28 dec, $^3J = 2.6$	–1.38 dec, $^4J = 0.1$	–51.09 t, $^3J = 0.8$